

Physicochemical And Preliminary Study on Gc-MS Analysis of *Prosopis africana* Seed (African Mesquite) Oil

Agubosi, O.C.P

Oluwafemi, R.A

Alagbe, J.O*

Department of Animal Science, University of Abuja, Gwagwalada, Nigeria

*Corresponding Author: Alagbe, J.O

Email: dralagbe@finmail.com

Abstract: A preliminary study was carried out to examine the various bioactive compounds and physicochemical properties of *Prosopis africana* seed oil (*African mesquite*). The extracted oil was yellow in colour with a yield of 10.73 % at liquid state, acid value, saponification, iodine, peroxide values ranged between 15.18 (mg NaOH/g of oil), 112.7 (mg/KOH/g of oil), 150.4 (gI₂/100g of oil) and 27.75 (meq/kg) respectively. A total number of 24 bioactive compounds was identified using GC-MS this includes: 2-phenyl propane (3.09 %), 1,3,5-trimethyl benzene (4.80 %), prosogerin A (12.67 %), prosogerin B (10.06 %), prosogerin C (7.11 %), prosogerin D (4.05 %), prosogerin E (2.89 %), luteolin 7-O-β-D-glucoside (0.67 %), quercetin 3-O-rutinoside (4.88 %), isoprosopilosine (4.28 %), tryptamine (1.66 %), β-phenethylamine (5.63 %), gallic acid (0.57 %), 4-hydroxy benzoic acid (1.41 %), 2,4-bis (1-phenylethyl) phenol (9.60 %), 2,4,6-tris(1-phenylethyl) phenol (7.83 %), rhodoxanthin (1.67 %), ferulic acid (1.20 %), pheophytin A (1.54 %), pheophytin B (3.31 %), campesterol (2.80 %) and β-sitosterol (1.51 %). Prosogerins are the most abundant compounds in the sample and are known to possess antimicrobial, anti-inflammatory and capable of scavenging free radicals thus preventing diseases in animals. It was concluded that *Prosopis africana* seed oil can be used as potential alternative to antibiotics and can be used to bridge the gap between food safety and livestock production.

Key words: *Prosopis africana*, seed, oil, phytochemicals, antimicrobials, antioxidants, free radicals

Introduction

Seeds from plants are potential reservoirs for bioactive compounds or secondary metabolites, proteins, carbohydrates, amino acid and fats. Recently, there has been increasing awareness on food safety and the use of plants as potential alternatives to antibiotics due to antimicrobial resistance, toxic residues of synthetic drugs in livestock products, increase in different ailments and death (Singh *et al.*, 2021). Plants of medicinal origin contains essential oils, which have been proven to be rich in phytochemicals (tannins, flavonoids, phenols, saponins, alkaloids, terpenoids etc), effective and generally regarded as safe (Adewale *et al.*, 2021). The high cost of animal protein has directed interest towards several seed protein as potential sources of vegetable protein for human food due to its nutrient compositions (Musbau and Asiru, 2020).

Prosopis africana (*African mesquite*) is a tropical leguminous plant. It belongs to the family Fabaceae. The tree reaches about 4-20 m in height; has an open crown and slightly rounded buttresses; bark is very dark, scaly, slash, orange to red-brown with white streaks (Aremu *et al.*, 2007). Seeds from *Prosopis africana* are used to prepare traditional fermented soup condiment or as flavor enhancers in Northern parts of Nigeria (Tajudeen *et al.*, 2011). The seeds are rich in several bioactive compounds which makes a medicinal plant (Oloyede, 2005). The ripe pods of *P. africana* are harvested by shaking off the ripe pod from the tree branches. According to Musbau and Asiru (2020) proximate composition of fermented *P. africana* seeds and pods revealed the presence of crude protein (27.75 %, 7.62 %), carbohydrate (27.09 %, 72.72 %), ether extracts (10.52 %, 6.86 %), fibre (6.76 %, 3.28 %), moisture (0.03 %, 5.62 %) and ash (6.04 %, 3.90 %). Oluwafemi *et al.* (2021) also reported that *P. africana* seed oil contains crude protein, crude fibre, ether extract, ash and energy at 30.71 %, 6.47 %, 3.66 %, 5.08 % and 383.26 (Kcal/kg).

The pods are useful in the feeding of ruminant animals due to their high nutritive value and can also be used traditionally for the treatment of tooth ache and other skin infections (Olorunmaiye *et al.*, 2019). Physicochemical property is an index used to evaluate the edibility as well as industrial application of oil.

This experiment was designed to examine the physicochemical properties as well as the secondary metabolites in *Prosopis africana* seed oil using GC-MS. This research will give a clue on its medicinal properties as well as the physiological actions of the oil in the body of animals

Materials and methods

Experimental site

This study was carried out at the department of Animal Science Research Laboratory, University of Abuja, along airport road, Gwagwalada, Abuja, Nigeria. Gwagwalada is located between latitudes 8°57¹ and 8°55¹N and longitude 7°05¹ and 7°06¹E.

Extraction of *Prosopis africana* seed

Mature seeds from *Prosopis africana* were harvested from different trees at the University of Abuja, Teaching and Research Farm. It was identified and authenticated by a certified taxonomist at the Department of Biological Sciences, University of Abuja, Gwagwalada with a voucher specimen number ANS/08F/2020. The seeds were sorted to remove the bad ones and washed with running water to remove dirt's and air dried for 18 days to maintain the bioactive chemicals in the seeds, mechanically separated from their pods with knife to obtain the seeds. Dried seeds was grinded using a laboratory grinder (Panasonic: model AS/309F) to obtain *Prosopis africana* seed meal and stored in a clean well labeled container for analysis. Prior to the commencement of the analysis, laboratory equipments were serviced and all the necessary reagents were purchased.

Extraction of PASO was done using a cold press machine (Model: ZZYX168, China). The grinded *Prosopis africana* (2000 g) was poured at into the feeder of the machine which works at a low temperature and PASO was collected via the squeeze cage.

Gas chromatography – Mass spectrometry analysis (GC-MS)

GC-MS analysis were performed on a GC – 2010 Shimadzu capillary gas chromatography directly coupled to the mass spectrometer system (GC-MS – model QP 2010; S/N column (70464300019 SA; Shimadzu) DB – 5ms non polar fused silica capillary column (30m X 0.25mm, 0.25µm film thickness) was used under following conditions: oven temperature program isotherm 2 min at 70°C, 3°C/min gradient to 200°C and final temperature kept for 35 min; injection temperature 200°C carrier gas is helium with flow rate 1.51ml/min; linear velocity 45.1 cm/sec. The effluent of the GC column was introduced directly into the source of MS and spectra obtained in the EI mode with ionization energy 70eV, in the electronic ionization mode and ion source temperature is 200°C. The solvent cut time 3 min. The sector mass analyzer was set to scan from 40 to 1000m/z with interface temperature of 240°C. The components of the essential oil were identified on the basis of comparison of their relative indices and mass spectra by computer matching with National Institute of Standards and Technology (NIST08) libraries.

Physicochemical analysis of *Prosopis africana* oil

Determination of acid value, saponification value, iodine value and acid values were determined according to the methods outlined by AOAC (2005).

Gas chromatography mass spectrophotometer (GC-MS) composition of *Prosopis africana* oil

The GC-MS result of *Prosopis africana* seed oil revealed many bioactive compounds. These compounds with their molecular weight, molecular formulae, retention time and their composition (%) determined from their peak areas is presented in Table 1. A total number of 24 bioactive compounds was identified including: 2-phenyl propane (3.09 %), 1,3,5-trimethyl benzene (4.80 %), prosogerin A (12.67 %), prosogerin B (10.06 %), prosogerin C (7.11 %), prosogerin D (4.05 %), prosogerin E (2.89 %), luteolin 7-O-β-D-glucoside (0.67 %), quercetin 3-O-rutinoside (4.88 %), isoprosopilosine (4.28 %), tryptamine (1.66 %), β-phenethylamine (5.63 %), gallic acid (0.57 %), 4-hydroxy benzoic acid (1.41 %), 2,4-bis (1-phenylethyl)

phenol (9.60 %), 2,4,6-tris(1-phenylethyl) phenol (7.83 %), rhodoxanthin (1.67 %), ferulic acid (1.20 %), pheophytin A (1.54 %), pheophytin B (3.31 %), campesterol (2.80 %) and β -sitosterol (1.51 %). The result obtained in this study agrees with the report of Peter *et al.* (2009); Ferguson *et al.* (2005); Bhardwaj *et al.* (1979, 1981); Simpson and Solbrig (1977). Bioactive compounds in plants are also regarded as phytochemicals which have been proven to be relatively cheap, safe effective and recently projected as suitable alternative to antibiotics (Adewale *et al.*, 2021; Musa *et al.*, 2020). For instance, prosogerins are group of flavonoids which performs multiple pharmacological functions such as: antibacterial, anti-inflammatory, antiviral, antitumor, hypolipidaemic, antiprotozoal, cytoprotective, antioxidant, antiplatelet and so on (Harzallah and Jannet, 2005; Alagbe and Motunrade, 2019; Alagbe, 2019, 2018). Prosogerins (A, B, C, D and E) are the most abundant compounds in this study; this confirms the earlier findings of Valli *et al.* (2014). Isoprosopilosine, tryptamine and β -phenethylamine are group of alkaloids which have diverse physiological effects: antibacterial, antimitotic, anti-inflammatory, analgesic, local anesthetic, hypnotic, psychotropic, and antitumor activity and many others (Aneela *et al.*, 2014; Ukani *et al.*, 2000). Alkaloids are also capable of working as an antimicrobial, antispasmodic and scavenging free radicals to prevent disease in the body of animals (Alagbe and Motunrade, 2019; Latif *et al.*, 2003). Studies have also reported the beneficial effects of phenol as an antioxidant, therefore the presence of 2,4-bis (1-phenylethyl) phenol and 2,4,6-tris(1-phenylethyl) phenols is a clear indication that *Prosopis africana* seed oil can function as an antibacterial (Kasolo *et al.*, 2010; Singh *et al.*, 2021) and can be traditionally used to treat skin diseases, wounds and other ailments.

Various bioactive chemicals in *Prosopis africana* seed oil exhibit a wide spectrum of antibacterial activities against pathogenic organisms thus influencing voluntary intake in animals and bile secretion (Zeng *et al.*, 2015). Composition of *Prosopis africana* seed oil varies according to plant species, geographical origin, season, environmental factors, processing techniques and storage conditions (Gadde *et al.*, 2017).

Table 1 Major chemical compounds of *Prosopis africana* oil as detected by gas chromatography mass spectrophotometer (GC-MS)

Chemical compounds	RI	% Area	MW(g/mol)	MF	Mass peak
2-phenyl propane	796	3.09	136.2	C ₉ H ₁₂ O	27, 42, 54, 79
1,3,5-trimethyl benzene	951	4.80	120.2	C ₉ H ₁₂	27, 55, 101, 134
Prosogerin A	1224	12.67	312.3	C ₁₇ H ₁₂ O ₆	27, 87, 102, 141
Prosogerin B	1009	10.06	330.8	C ₁₈ H ₁₄ O ₇	27, 45, 58, 105
Prosogerin C	806	7.11	372.4	C ₂₀ H ₂₀ O ₇	27, 62, 71, 90, 166
Prosogerin D	1205	4.05	358.3	C ₁₉ H ₁₈ O ₇	27, 41, 48, 54, 71
Prosogerin E	1104	2.89	344.3	C ₁₈ H ₁₆ O ₇	27, 38, 51, 74, 89
Luteolin 7-O- β -D-glucoside	1967	0.67	447.4	C ₂₁ H ₁₉ O ₁₁	27, 38, 77, 96, 155
Quercetin 3-O-rutinoside	2196	4.88	610.51	C ₂₇ H ₃₀ O ₁₆	27, 63, 89, 94, 106
Apigenin-8-glucoside	1663	3.31s	564.5	C ₂₆ H ₂₈ O ₁₄	27, 47, 69, 76, 93
Quercetin-3-glucoside	1907	5.43	464.09	C ₂₁ H ₂₀ O ₁₂	27, 85, 89, 94, 97
Isoprosopilosine	1800	4.28	933.4	C ₅₇ H ₁₀₄ O ₉	27, 41, 55, 81, 95
Tryptamine	2506	1.66	160.2	C ₁₀ H ₁₂ N ₂	27, 41, 57, 93, 121
β -phenethylamine	1002	5.63	121.18	C ₈ H ₁₁ N	27, 67, 80, 89, 93
Gallic acid	2707	0.57	170.12	C ₇ H ₆ O ₅	27, 88, 89, 97, 109
4-hydroxy benzoic acid	779	1.41	138.12	C ₇ H ₆ O ₃	27, 40, 43, 86, 92
2,4-bis (1-phenylethyl) phenol	1117	9.60	330.5	C ₂₄ H ₂₆ O	27, 45, 57, 66, 72
2,4,6-tris(1-phenylethyl) phenol	1251	7.83	406.6	C ₃₀ H ₃₀ O	27, 48, 53, 67

Rhodoxanthin	2265	1.67	562.9	C ₄₀ H ₅₀ O ₂	27, 40, 43, 48, 60
Ferulic acid	1307	1.20	194.18	C ₁₀ H ₁₀ O ₄	27, 38, 40, 54
Pheophytin A	922	1.54	871.2	C ₅₅ H ₇₄ N ₄ O ₅	27, 42, 47, 76, 78
Pheophytin B	1006	3.31	885.2	C ₅₅ H ₇₂ N ₄ O ₆	27, 34, 37, 38
Campsterol	1227	2.80	400.7	C ₂₈ H ₄₈ O	27, 32, 43, 47
β-sitosterol	2093	1.51	414.71	C ₂₉ H ₅₀ O	27, 45, 30, 45, 52

RI: retention index; MW: molecular weight; MF: molecular formula

Physicochemical composition of *Prosopis africana* seed oil

Physicochemical composition of *Prosopis africana* seed oil is presented in Table 2. The extracted oil was yellow in colour with a yield of 10.73 % at liquid state. Acid, saponification, iodine, peroxide values ranged between 15.18 (mg NaOH/g of oil), 112.7 (mg/KOH/g of oil), 150.4 (gI₂/100g of oil) and 27.75 (meq/kg) respectively. The physical evaluation of *Prosopis africana* seed oil shows that the colour of the oil was light yellow and it has no offensive odour. However, the oil yield was lower than 18 % reported for soya bean seed and 43 % for groundnut seed (Franz *et al.*, 2011). They are however higher than 1.42 % reported for the seeds of *Piliostigma thonningii* (Deepak *et al.*, 2012). Refractive index is the ratio of the velocity of light in vacuum to the velocity of light in the medium is an indication of the level of saturation of the oil (Isman *et al.*, 2007). Iodine value is a measure of degree of unsaturation and it is an identity characteristic of seed oils, making it an excellent raw materials for soaps and cosmetic industries (Joulang and Konig, 1998). Acid value gives an indication of the quantity of free fatty acids in oil.

Saponification value is a measure of oxidation during storage and also indicates deterioration of the oils (Hedges and Lister, 2006). Peroxide value is a measure of peroxides contained in the oil and used in determining the degree of spoilage. The standard peroxide value for edible oils which have not undergone rancidity must be well below 10 meq/kg (Olayemi *et al.*, 2018).

Table 2 Physicochemical composition of *Prosopis africana* seed oil

Parameter	Composition	*WHO (edible oil)
Colour	Yellow	NS
Odour	Agreeable	NS
State	Liquid	-
% Yield	10.73	NS
Acid value (mg NaOH/g of oil)	15.18	3.80 – 4.00
Saponification value (mg/KOH/g of oil)	112.7	170.0 – 181.0
Iodine value (gI ₂ /100g of oil)	150.4	100.0 – 150.0
Peroxide value (meq/Kg)	27.75	10.0
Refractive index at (20°C)	2.01	1.47 – 1.51
Specific gravity at (25°C)	1.02	0.80 – 0.84
% Free fatty acid	14.10	5.78 -7.28

*WHO (2007)

Conclusion

Prosopis africana seed oil contains chemical compounds with recognized medicinal value/therapeutics, or which are precursors for chemo-pharmaceutical semi-synthesis. The medicinal value of this oil lies in some chemical substances (phytochemicals or bioactive chemicals) that produce a definite physiological action in the body of animals. The major bioactive compounds in *Prosopis africana* seed oil are prosogerins and their composition and concentration vary according to the plant, parts of the plant, geographical origin, harvesting season, environmental factors, storage conditions, and processing techniques.

References

1. Kasolo, J.N., Gabriel, S., Bimenya, L.O., Joseph, O. and Ogwal-Okeng, J.W. (2010). Phytochemicals and uses of Moringa oleifera leaves in Ugandan rural communities. *J. Med. Plants Res.* 4(9): 753-757.
2. Latif A, Ahmad H, Begum S, Adnan M, Hassian S, Waseem M. (2003). Medicinal and other economic plants as substitute to forest logging in Miandam and Sulatanr valleys, Swat. Proceedings of international workshop on conservation and sustainable use of medicinal and aromatic plants in Pakistan. WWF Pak., 101-105
3. Gadde, U., Kim, W.H., Oh, S.T and Lillehoj, H.S. (2017). Alternatives to antibiotics for maximizing growth and feed efficiency in Poultry: A review. *Journal of Animal Science Research.* 18: 26-45.
4. Musa, B., Alagbe, J.O., Adegbite Motunrade Betty, Omokore, E.A. (2020). Growth performance, caeca microbial population and immune response of broiler chicks fed aqueous extract of *Balanites aegyptiaca* and *Alchornea cordifolia* stem bark mixture. *United Journal for Research and Technology*, 2(2):13-21.
5. Alagbe, J.O and Adegbite Motunrade Betty (2019). Haematological and serum biochemical indices of starter broiler chicks fed aqueous extract of *Balanites aegyptiaca* and *Alchornea cordifolia* bark mixture. *International Journal of Biological, Physical and Chemical Studies.* 1(1): 8-15
6. Alagbe, J.O (2020). Proximate, phytochemical and vitamin compositions of *Prosopis africana* stem bark. *European Journal of Agricultural and Rural Education.* 1(4): 1-7.
7. Oluwafemi, R.A., Akinbisola, S.A and Alagbe, J.O. (2020). Nutritional and growth performance of feeding *Polylathia longifolia* Leaf Meal as partial replacement of Wheat Offal in the diet of broiler chicks. *European Journal of Biotechnology and Bioscience.* 8(4): 17-21.
8. Singh, A.S., Alagbe, J.O., Sharma, S., Oluwafemi, R.A and Agubosi, O.C.P. (2021). Effect of dietary supplementation of melon (*Citrullus linatus*) seed oil on the growth performance and antioxidant status of growing rabbits. *Journal of Multidimensional Research and Reviews*, 2(1): 78-95.
9. Alagbe, J.O. (2021). *Prosopis africana* stem bark as an alternative to antibiotic feed additives in broiler chicks diets: Performance and Carcass characteristics. *Journal of Multidimensional Research and Reviews*, 2(1): 64-77.
10. Adewale, A.O., Alagbe, J.O., Adeoye, Adekemi. O. (2021). Dietary Supplementation of *Rauvolfia Vomitoria* Root Extract as A Phytogenic Feed Additive in Growing Rabbit Diets: Haematology and serum biochemical indices. *International Journal of Orange Technologies*, 3(3): 1-12.
11. Oloyede, I.O. (2005). Chemical profile of Unripe pulp of Carica papaya. *Pakistan Journal of Nutrition* 6: 379-381.
12. Tajudeen, A.I., Kayode, C.O., Yahaya, A and Mohammed, S.A. (2011). Development and testing of a *Prosopis* pod thresher. *Australian Journal of Basic and Applied Sciences* 5(2): 759-767.
13. Olorumaiye, K.S., Apeh, L.E., Madandola, H.A and Oguntoye, M.O. (2019). Proximate and phytochemical composition of African mahogany seed and *African mesquite* pod. *Journal of Applied Sciences and Environmental Management* 23(2): 249-252.
14. Musbau, S and Asiru, R.A. (2020). Proximate parameters of fermented *Prosopis africana* seeds. *Journal of Academia and Industrial Research* 8(9): 163-165.
15. Aremu, M.O., Olanisakin, B.O., Atolaye, B.O and Ogbu, C.F. (2007). Some nutritional composition and functional properties of *Prosopis africana*. *Bangladesh Journal of Industrial Research* 4(3): 269-270.
16. Aneela, S., Dey, A and De, S. (2014). GC-MS analysis of methanolic extract of *Prosopis spicigera*. *International Journal of Phytopharmacology* 5: 168-171.
17. Ukani, M.D., Limbani, N.B and Mehta, N.K. (2000). A review on the Ayurvedic herb *Prosopis cineraria* (L) Druce. *Anc. Sci. Life* 20:58-70
18. Simpson, B.B and Solbrig, O.T. (1977). Introduction Mesquite: its biology in two desert ecosystems. Inc. Stroudsberg, pp 1-7.
19. Peter, S., Stephene, D., Eric, M and Phillipe, G. (2009). Unusal amount of (-) mesquitol from the heartwood of *Prosopis juliflora*. *Nat. Prod. Res.* 23: 183-189.

20. Valli, S., Gokulshankar, S., Mohanthy, B.K., Ranjith, M.S., Ashutosh, S.R and Remya, V. (2014). Anticryptococcal activity of alkaloid rich fraction of leaves of *Prosopis juliflora*. A future promising supplementary therapy for cryptococcosis and cyyptococcal meningitis. Int. J. Pharm. Sci. 6: 491-495.
21. Harzallah, S.F and Jannet, H.B. (2005). Flavonoids diversification in organs of two *Prosopis fructa* (Bank and Sol.) eig. (Leguminosea, Mimosoidae) populations occurring in Northeast and the Southeast of Tunisia. Journal of Applied Research 1: 130-136.
22. Ferguson, L.R., Shuo-tun, Z and Harris, P. (2005). Antioxidant and antigenotoxic effects of plant cell wall hydroxycinnamic acids in cultured HT-29 cells. Molecular Nutrition and Food Research 49: 585-593.
23. Bhardwaj, D.K., Bisht, M.S., Mehta, C.K and Sharma, G.C. (1979). Flavonoids of *Prosopis spicigera* flowers. Journal of Phytochemistry 18: 355-356.
24. Bhardwaj, D.K., Gupta, A.K., Jain, R.K and Sharma, G.C. (1981). Chemical examination of *Prosopis spicigera* seeds. J. Nat. Prod. 44: 656-659.
25. Oluwafemi, R.A., Agubosi, O.C.P and Alagbe, J.O. (2021). Proximate, minerals, vitamins and amino acid composition of prosopis africana (*African mesquite*) seed oil. Asian Journal of Advances in Research 11(1): 21-25.
26. Olayemi, R.F., Jawonisi, I.O and Samuel, J.A. (2018). Characterization and physiochemical analysis of essential oil of *Cymbopogon citratus* leaves. Bayero Journal of Pure and Applied Sciences. 11(1): 74-81.
27. Alagbe, J.O., Shittu, M.D., Bamigboye, S.O and Oluwatobi, O. (2019). Proximate and mineral analysis of *Pentadiplandra brazzeana* stem bark. Electronic Research Journal of Engineering and Applied Sciences 1(2019): 91-99.
28. Alagbe, J.O and Oluwafemi, R.A (2019). Growth performance of weaner rabbits fed noni and moringa olifera leaf as partial replacement for soya bean meal. International Journal of Advanced Biological and Biomedical Research. 7(2): 185-195.
29. Alagbe, J.O. (2018). Performance and haemato-biochemical parameters of weaner rabbits fed diets supplemented with dried water melon rinds meal. Journal of Dairy and Veterinary Sciences 8(4): 2018.
30. Deepak, G., Ashish, K.G and Ritam, M. (2012). An update on bioactive potential of a monoterpene aldehyde citral. Journal of Biologically Active Products from Nature 2(4): 186-199.
31. Franz, A.R., Knaak, N and Fiuza, L.M. (2011). Toxic effects of essential plant oils in adult *Sitophilus oryzae*. Revista Brasileira de Entomologia. 55: 116-120.
32. Hedges, L.J and Lister, C.E. (2006). Nutritional attributes of Brassica vegetables Crop and Food Confidential report no. 1618. New Zea Insti Crop & Food Res Ltd 46.
33. Isman, M., Machial, C., Miresmailli, S and Bainard, L. (2007). Essential oil based pesticides: new insights from old chemistry in: Ohkawa Wiley-VCH, Weinheim Germany pp. 201-209.
34. Joulang, D and Konig, W.A. (2018). The atlas of spectra data of sesquiterpene hydrocarbons Verlay, Hamburg, Germany pp. 115-123.