# Broiler Microbiological and Histological Response to Diets Varying in Sodium and Calcium Butyrate

Mayada Saeed Sultan Wasit Agriculture Directorate, Wasit, Iraq Email: mayada.said1201a@coagri.uobaghdad.edu.iq

### Abstract

The aim of the study was to determine the effect of supplementing broiler diets with different levels of sodium and calcium butyrate on certain histological and microbial traits. This study was conducted in the poultry field of the Department of Animal Production at the College of Agricultural Engineering Sciences / University of Baghdad (Baghdad, Iraq) during November - December (2020) in a total of 308 one-day-old chicken utilized in the study, and each treatment had three replicates consisting of 10 chicks each. Duodenal villi height increased significantly (p<0.05) for treatments T2, T3, T4, and T5, villi thickness increased significantly (p<0.05) for treatment T3, and crypt depth increased significantly (p<0.05) for treatment T2. Treatment T2 significantly increased villi height and treatment T5 significantly increased villi thickness in the jejunum, in comparison to the other treatments. In the ileum, treatment T2 resulted in significantly greater increases in villi height, treatment T6 in villi thickness, and treatment T4 in crypt depth compared to the other treatments. In the duodenum and jejunum, there were no statistically significant variations in the numbers of *E. coli* and Lactobacilli bacteria, but in the ileum, the number of E. coli increased significantly in treatments T3 and T4. In this respect, lactobacilli did not differ noticeably.

Keywords: Lactobacilli, Chicken, Ileum, Duodenum, Jejunum

### Introduction

Antibiotics and other additives have long been used in poultry feed as growth promoters; their addition has been shown to stabilize the intestinal microbial flora, improve growth performance, and prevent some intestinal diseases (Hassan et al., 2010). These are two requirements of the modern poultry industry (Javid and Khan, 2016). Because of rising antibiotic resistance in chickens (Kabir 2009) and the risk of spreading disease from infected animals to humans, the European Union banned the use of antibiotics in poultry feed in 2006 (Singer and Hofacre 2006; Tokić et al 2007), requiring the poultry industry to develop methods for raising birds without antibiotics (Castanon, 2007). Organic acids and their salts, like butyric acid, are a byproduct of microbial fermentation of dietary fibers (Hamer et al., 2008) and are considered natural products with no harmful effect; they can be used with all species and ages of birds and do not interact with most medicines and feed additives in bird feed or drinking water. They extend the life of feed and make it easier to store (Zhang et al., 2014; Coban, 2020). Since organic acids and their salts reduce the pH of the digestive tract, they have attracted a lot of attention for their potential to inhibit the growth of harmful bacteria (Eidelsburger et al., 1992; Boling et al., 2000; Partanen et al., 2001; Kill et al., 2011a). These compounds have also been shown to have beneficial effects on growth performance and intestinal integrity (Chamba et al., 2014) Na, K, Mg, and Ca salts of butyric acid can be utilized as food additives because, unlike free acids, butyric acid has no odor and is less volatile, making it safer to work with during the feed manufacturing process (Ahsan et al., 2016). In addition to improving intestinal health and preventing the emergence of cancer cells in the colon epithelial cells by regulating cell proliferation, differentiation, and apoptosis (Da Zhou et al. 2017), the salts have other benefits, such as stimulating intestinal blood flow, mucin secretion, and water absorption. The capacity of acid salt to inhibit the development of pathogenic bacteria has led to its use as a feed additive (Flint et al., 2012). The aim of the study was to determine the effect of different levels of sodium butyrate and calcium butyrate on broilers' histological and microbiological characteristics.

#### Materials and methods

From November 1, 2020, to November 12, 2020, the University of Baghdad's Department of Animal Production's poultry farm served as the site for a field experiment in the College of Agricultural Engineering Sciences. The experiment used 210 Ross-308 broiler chicks that were one days old and had not yet been sexed from the Iraqi Society hatchery in Abu Ghraib, Baghdad. Chicks typically weighed around 42,372 grammes on average. The chicks were given food and water ad libitum and were grown on a layer of sawdust that was 3 to 5 centimeters deep. The lighting was set up so that there was 23 hours of light and one hour of darkness to help the chicks adjust to the dark. Diets were prepared in a laboratory dedicated to the chicken industry, and chicks were randomly assigned to one of seven treatments, with three replicates per treatment (10 birds per replicate). Statistical analysis was carried out using One-Way ANOVA in the SPSS Software. Differences were considered significant at P<0.05 (Al-Gharban, 2017)

Ingradiants	Starter diet						
Ingreutents	<b>T1</b>	<b>T2</b>	<b>T3</b>	<b>T4</b>	T5	<b>T6</b>	<b>T7</b>
Yellow corn	43.8	43.8	43.6	43.4	43.5	43.5	43.5
Wheat	14	14	14	13.5	14	14	13.7
Soybean meal	32.7	32.5	32.5	32.6	32.7	32.7	32.7
Protein Center (1)	5	5	5	5	5	5	5
oil	2.2	2.2	2.2	2.4	2.4	2.3	2.3
<b>Di-Calcium Phosphite</b>	0.7	0.7	0.7	0.7	0.7	0.7	0.7
Limestone	1.1	1.1	1.1	1.1	1.1	1.1	1.1
Vitamins and Minerals Blend (2)	0.2	0.2	0.2	0.2	0.2	0.2	0.2
Calcium butyrate	-	-	-	-	0.2	0.4	0.8
Sodium butyrate	-	0.2	0.4	0.8	-	-	-
Salt	0.3	0.3	0.3	0.3	0.2	0.1	0.0
Total	100	100	100	100	100	100	100
Calculated Chemical Analysis (3)							
Crude protein %	23.0	22.9	22.9	22.8	23.0	23	22.9
Represented energy (Kg/ Kcal	3005 3	3000 /	2003 7	2001 8	3013.2	3004.2	2004 0
feed)	5005.5	3000.4	2993.1	2771.0	5015.2	3004.2	2774.7
Methionine + Cysteine %	0.88	0.88	0.88	0.88	0.88	0.88	0.88
Lysine %	1.3	1.3	1.3	1.3	1.3	1.3	1.3
Calcium %	0.88	0.88	0.88	0.88	0.88	0.88	0.88
Phosphorous %	0.48	0.48	0.48	0.48	0.48	0.48	0.48

Table 1.	Starter diet	(1-10  days)	) ingredients and	computed	chemical con	position
I UNIC II	Starter are	$(\mathbf{I} \mathbf{I} \mathbf{U} \mathbf{u} \mathbf{u} \mathbf{y} \mathbf{y})$	ingi cultito una	computed	chemical con	position

- (1)Dutch protein center BROCON-5 SPECIAL W. Each kg contains: 40% crude protein, 5% fat, 2.81% fiber, 3.14% calcium, 2.65% phosphorous, 2.50% sodium, 3.88% chloride, 3.85% lysine, 3.70% methionine, 4.12% methionine + cysteine, 2107 Kilo Calories/Kg Represented Energy, 20,000 IU Vitamin A, 80,000 IU Vitamin D3, 600 mg Vitamin E, 50 mg Vitamin K3, 50 mg Vitamin B1, 140 mg Vitamin B2, 80 mg Vitamin B6, 700 μg B12, 20 mg Acid Folic, 5 mg citric acid, 2 mg biotin, 800 mg niacin, 1 mg iron, 200 mg copper, 1,600 mg manganese, 1,200 mg zinc, 20 mg iodine, 5 mg selenium, 6 mg cobalt, 33.50 mg antioxidant (BHT).
- (2)A mixture of vitamins and minerals, each kg of which contains: 5000 IU Vitamin A, 600 IU D3, 10 mg E, 2 mg K3, 2 mg B1, 2 mg B2, 2 mg B6, 5 micrograms B12, 10 mg C, 15 mg niacin, 500 μg folic acid, 5 mg D-calcium phosphate, 40 mg zinc, 100 μg cobalt, 150 mg lysine.

(3)According to the chemical analysis of the ration according to the NRS (1994)

Table 2. Growth diet (11 – 22 days) ingredients and computed chemical co
--------------------------------------------------------------------------

Ingradianta	Growth diet						
Ingreulents	T1	T2	T3	T4	T5	<b>T6</b>	T7
Yellow corn	44.5	44.5	44.3	44	44.6	44.5	44.2
Wheat	15.9	15.7	15.7	15.7	15.7	15.7	15.5

# Texas Journal of Agriculture and Biological Sciences <u>https://zienjournals.com</u>

Soybean meal	29	29	29	28.5	29	29	29
Protein Center (1)	5	5	5	5	5	5	5
oil	3.4	3.4	3.4	3.4	3.4	3.4	3.6
Di-Calcium Phosphite	0.6	0.6	0.6	0.6	0.6	0.6	0.6
Limestone	1.1	1.1	1.1	1.1	1.1	1.1	1.1
Vitamins and Minerals Blend (2)	0.2	0.2	0.2	0.2	0.2	0.2	0.2
Calcium butyrate	-	-	-	-	0.2	0.4	0.8
Sodium butyrate	-	0.2	0.4	0.8	-	-	-
Salt	0.3	0.3	0.3	0.3	0.2	0.1	0.0
Total	100	100	100	100	100	100	100
Calculated Chemical Analysis (3)							
Crude protein %	21.5	21.5	21.4	21.2	21.5	21.5	21.4
Represented energy (Kg/ Kcal feed)	3105.5	3099.1	3092.4	3106.2	3102.5	3099.1	3100.9
Methionine + Cysteine %	0.84	0.84	0.82	0.83	0.84	0.84	0.83
Lysine %	1.2	1.2	1.2	1.2	1.2	1.2	1.2
Calcium %	0.85	0.85	0.85	0.85	0.85	0.85	0.85
Phosphorous %	0.46	0.46	0.46	0.46	0.46	0.46	0.46

(1)Dutch protein center BROCON-5 SPECIAL W. Each kg contains: 40% crude protein, 5% fat, 2.81% fiber, 3.14% calcium, 2.65% phosphorous, 2.50% sodium, 3.88% chloride, 3.85% lysine, 3.70% methionine, 4.12% methionine + cysteine, 2107 Kilo Calories/Kg Represented Energy, 20,000 IU Vitamin A, 80,000 IU Vitamin D3, 600 mg Vitamin E, 50 mg Vitamin K3, 50 mg Vitamin B1, 140 mg Vitamin B2, 80 mg Vitamin B6, 700 μg B12, 20 mg Acid Folic, 5 mg citric acid, 2 mg biotin, 800 mg niacin, 1 mg iron, 200 mg copper, 1,600 mg manganese, 1,200 mg zinc, 20 mg iodine, 5 mg selenium, 6 mg cobalt, 33.50 mg antioxidant (BHT).

(2)A mixture of vitamins and minerals, each kg of which contains: 5000 IU Vitamin A, 600 IU D3, 10 mg E, 2 mg K3, 2 mg B1, 2 mg B2, 2 mg B6, 5 micrograms B12, 10 mg C, 15 mg niacin, 500 μg folic acid, 5 mg D-calcium phosphate, 40 mg zinc, 100 μg cobalt, 150 mg lysine.

(3)According to the chemical analysis of the ration according to the NRS (1994).

	Table 3. Growth	diet (23 - 4	2 days) ingredients	and computed	chemical composition.
--	-----------------	--------------	---------------------	--------------	-----------------------

Ingradiants	Final diet						
Ingreutents	T1	T2	<b>T3</b>	T4	T5	<b>T6</b>	<b>T7</b>
Yellow corn	47.6	47.6	47.3	47.1	47.4	47.2	47.2
Wheat	15	14.8	14.8	14.2	15	15	14.5
Soybean meal	26	26	26	26.2	26	26	26.2
Protein Center (1)	5	5	5	5	5	5	5
oil	4.2	4.2	4.3	4.5	4.3	4.3	4.4
Di-Calcium Phosphite	0.6	0.6	0.6	0.6	0.6	0.6	0.6
Limestone	1.1	1.1	1.1	1.1	1.1	1.1	1.1
Vitamins and Minerals Blend (2)	0.2	0.2	0.2	0.2	0.2	0.2	0.2
Calcium butyrate	-	-	-	-	0.2	0.4	0.8
Sodium butyrate	-	0.2	0.4	0.8	-	-	-
Salt	0.3	0.3	0.3	0.3	0.2	0.1	0.0
Total	100	100	100	100	100	100	100
Calculated Chemical Analysis (3)							
Crude protein %	20.2	20.2	20.2	20.2	20.2	20.2	20.2
Represented energy (Kg/ Kcal feed)	3180.0	3173.7	3172.7	3170.1	3182.3	3184.6	3173.8
Methionine + Cysteine %	0.80	0.80	0.80	0.80	0.80	0.80	0.80
Lysine %	1.1	1.1	1.1	1.1	1.1	1.1	1.1
Calcium %	0.84	0.84	0.84	0.84	0.84	0.84	0.84
Phosphorous %	0.45	0.45	0.45	0.45	0.45	0.45	0.45

- (1)Dutch protein center BROCON-5 SPECIAL W. Each kg contains: 40% crude protein, 5% fat, 2.81% fiber, 3.14% calcium, 2.65% phosphorous, 2.50% sodium, 3.88% chloride, 3.85% lysine, 3.70% methionine, 4.12% methionine + cysteine, 2107 Kilo Calories/Kg Represented Energy, 20,000 IU Vitamin A, 80,000 IU Vitamin D3, 600 mg Vitamin E, 50 mg Vitamin K3, 50 mg Vitamin B1, 140 mg Vitamin B2, 80 mg Vitamin B6, 700 μg B12, 20 mg Acid Folic, 5 mg citric acid, 2 mg biotin, 800 mg niacin, 1 mg iron, 200 mg copper, 1,600 mg manganese, 1,200 mg zinc, 20 mg iodine, 5 mg selenium, 6 mg cobalt, 33.50 mg antioxidant (BHT).
- (2)A mixture of vitamins and minerals, each kg of which contains: 5000 IU Vitamin A, 600 IU D3, 10 mg E, 2 mg K3, 2 mg B1, 2 mg B2, 2 mg B6, 5 micrograms B12, 10 mg C, 15 mg niacin, 500 μg folic acid, 5 mg D-calcium phosphate, 40 mg zinc, 100 μg cobalt, 150 mg lysine.
- (3)According to the chemical analysis of the ration according to the NRS (1994).

## **Results and discussion**

### Histological traits

According to Table (4), adding different level of sodium and calcium butyrate to broiler diets from 1 to 42 days old led to a statistically significant (P0.05) rise in duodenal villi height for treatments T2, T3, T4, and T5 compared with treatments T1, T6, and T7.Treatment T3 had the greatest villi thickness compared to the other experimental treatments, while treatment T2 had the greatest significant depth of crypts.

Fable 4. Effect of sodium and calcium butyrate supplementation on the histological characteristics o	f
the broiler duodenum at 6 weeks of age	

Tucotmonto	Duodenum					
Treatments	Villi height (µm)	Villi thickness (µm)	Crypt depth (µm)			
<b>T1</b>	C 2.66 ±175.94	B 2.266 ±48.02	BC 3.34 ±57.92			
T2	A 8.88 ±338.91	B 3.25 ±51.42	A 2.54 ±73.08			
T3	A 8.28 ±340.66	A 1.54 ±58.64	B 2.33 ±61.55			
T4	A 4.80 ±335.85	B 2.00 ±49.49	C 2.22 ±54.13			
T5	A 9.08 ±317.20	B 1.46 ±50.69	BC 2.00 ±59.24			
<b>T6</b>	C 3.01 ±187.60	B 1.70 ±47.09	BC 1.05 ±58.94			
<b>T7</b>	B11.43 ±244.43	B 1.21 ±45.97	BC 1.39 ±55.66			
Leave of significant	*	*	*			

\*Different letters within the same column indicate significant differences at the level (P<0.05), N.S. There is no significant difference.

\*\* treatments: T1 is a control treatment free of addition, T2, T3, T4, adding sodium butyrate at levels 0.2, 0.4, 0.8%, respectively, and T5, T6, and T7 adding calcium butyrate at levels 0.2, 0.4, 0.8%, respectively.

Treatment T2 showed a significant increase (P<0.05) in jejunal villi height compared to the other experimental treatments, while treatment T5 showed a significant increase in villi thickness compared to the other treatments. The depth of the crypts significantly increased with the treatments T1, T2, and T5 (table 5). **Table 5. The effect of sodium and calcium butyrate supplementation on the histological characteristics of the broiler Jejunum at 6 weeks of age** 

Treatmonts	Jejunum						
Treatments	Villi height (µm)	Villi thickness (µm)	Crypt depth (µm)				
T1	AB 9.41 ±330.39	BC 2.41 ±52.28	A 2.91 ±74.18				
T2	A 9.35 ±359.91	AB 2.52 ±57.63	A 7.73 ±72.08				
T3	E 4.45 ±227.41	D 3.06 ±42.62	BC 1.77 ±53.60				
T4	CD 11.22 ±276.93	CD 1.39 ±45.26	C 2.27 ±46.13				
T5	BC 7.27 ±302.08	A 2.78 ±61.26	A 1.60 ±74.62				
T6	CD 12.25 ±272.37	BC 1.17 ±52.49	BC 2.27 ±56.24				
T7	DE 9.10 ±253.06	BC 1.71 ±50.48	B 1.93 ±57.11				
Leave of significant	*	*	*				

\*Different letters within the same column indicate significant differences at the level (P<0.05), N.S. There is no significant difference.

\*\* treatments: T1 is a control treatment free of addition, T2, T3, T4, adding sodium butyrate at levels 0.2, 0.4, 0.8%, respectively, and T5, T6, and T7 adding calcium butyrate at levels 0.2, 0.4, 0.8%, respectively.

Treatment T2 showed a significant increase (P<0.05) in villi compared to the other treatments, while treatment T6 showed a significantly higher level of villous thickness and treatment T4 showed a significantly higher level of crypt depth (Table 6).

Table 6. The effect of sodium and calcium butyrate supplementation on the histological characteristics			
of the broiler ileum at 6 weeks of age			

Treatmonts	lleum						
Treatments	Villi height (µm)	Villi thickness (µm)	Crypt depth (µm)				
T1	B 8.07 ±340.15	ABC 2.59 ±49.65	C 3.59 ± 58.39				
T2	A 11.20 ±406.33	ABC 1.84 ±49.87	C 2.35 ± 74.5				
T3	BC 9.61±324.27	AB 2.62 ±53.87	C 2.91 ±53.47				
T4	BC 10.90 ±323.31	BC 2.56 ±47.38	A 10.90 ±323.31				
T5	C 7.20 ±294.27	C 2.38 ±45.47	B 29.68 ±136.69				
T6	D 10.45 ± 238.90	A 1.80 ±55.61	$C 3.52 \pm 53.07$				
T7	D 11.05 ±229.16	ABC1.45 ±52.43	C 1.42 ±55.76				
Leave of significant	*	*	*				

\*Different letters within the same column indicate significant differences at the level (P<0.05), N.S. There is no significant difference.

\*\* treatments: T1 is a control treatment free of addition, T2, T3, T4, adding sodium butyrate at levels 0.2, 0.4, 0.8%, respectively, and T5, T6, and T7 adding calcium butyrate at levels 0.2, 0.4, 0.8%, respectively.

Butyrate is a source of energy for the intestinal cells and thus contributes to the growth and development of the villi, which may explain why adding sodium and calcium butyrate causes changes in the histological characterization (Lesson et al., 2005; Ahsan et al., 2016). Enterocyte proliferation and villus elongation were promoted by calcium butyrate in broiler diets, as reported by Yang et al. (2009) and Abd El Wahab et al. (2019).

The majority of the energy and physiological requirements for intestinal cells are met by short-chain fatty acids (Nagy, 2007), so supplementing with salts of these acids, such as sodium and calcium butyrate, may promote the development of goblet cells and mucin production, as well as the height of villi, the thickness of crypts, and the depth of crypts (Martinez et al., 2016). In addition to stimulating the development of goblet cells and mucin protected by butyric acid, which also enhances their histological properties and proliferation. The histological features of the jejunum and duodenum are enhanced by sodium and calcium butyrate. Increased differentiation of epithelial cells enhanced intestinal epithelial function, and preservation of villus structure have all been linked to mucosal tensing and protection (Hu et al., 2007; Al Fataftah and Abdelqader, 2016). This explains for the increased villi height, width, and crypt depth, as well as the encouragement of goblet cell proliferation and mucin production (Martinez et al., 2016).

Butyric acid protects the mucous membrane, promotes intestinal growth, and enhances the histological characteristics of the intestinal epithelium, just as sodium and calcium butyrate do for the jejunum and duodenum (Hu et al., 2007; Al Fataftah and Abdelqader, 2016). In addition to enhancing epithelial cell differentiation, enhancing epithelial cell function, and protecting villus structure (Kinoshita et al. 2002; Elnesr et al. 2019).Butyric acid also promotes intestinal cell proliferation, which aids in the repair of injured mucosa and increases villus height, as well as improves blood flow to the intestines and the secretion of digestive enzymes (Elnesr et al. 2017b; Sikandar et al., 2018).

Table 7 reveals that the number of E. coli was significantly higher in treatments T3, T4, T6, and T7 compared to T1, whereas there were no statistically significant differences between treatments T2, T5, and T1. As the pH of the intestine is lowered by calcium and sodium butyrate, it becomes an unfavourable environment for the growth of harmful bacteria and the activity of acid-loving bacteria (Hodin, 2000; Rezaei, and Guilloteau, 2010). The composition of the intestinal microbiota is profoundly influenced by acidity (Warnecke and Gill, 2005).

(regulationine cycle , g) in anter one parts of the interstine at a weeks of age						
Treatment	Duodenum		Jejunum		Ileum	
	E. coli	Lacto	E. coli	Lacto	E. coli	Lacto
T1	0.11±4.74	0.18±4.99	0.13±4.87	0.08±4.93	C 0.16±4.73	0.07±5.18
T2	0.06±4.88	0.02±5.27	0.50±5.22	0.29±4.85	BC 0.05±4.92	0.45±5.56
T3	0.31±5.33	0.20±4.86	0.45±5.19	0.29±4.68	A 0.05±5.24	0.20±5.06
T4	0.07±5.19	0.27±4.72	0.01±4.90	0.12±5.06	AB 0.09±5.12	0.04±5.26
T5	0.11±4.95	0.32±5.25	0.09±494	0.32±5.43	C 0.13±4.78C	0.32±5.47
T6	0.37±5.42	0.12±5.20	0.23±5.10	0.47±5.72	BC 0.02±4.90	0.22±5.11
T7	0.26±5.40	0.08±5.16	0.36±5.50	0.61±4.92	BC 0.10±4.90	0.39±5.42
Significance	N.S	N.S	N.S	N.S	*	N.S

 Table 7. Effect of sodium and calcium butyrate supplementation on the on microbial traits (logarithmic cycle / g) in different parts of the intestine at 6 weeks of age

\*Different letters within the same column indicate significant differences at the level (P<0.05), N.S. There is no significant difference.

\*\* treatments: T1 is a control treatment free of addition, T2, T3, T4, adding sodium butyrate at levels 0.2, 0.4, 0.8%, respectively, and T5, T6, and T7 adding calcium butyrate at levels 0.2, 0.4, 0.8%, respectively.

### Reference

- 1. Abd El-Wahab, A., Mahmoud, R. E., Ahmed, M. F., and Salama, M. F. (2019). Effect of dietary supplementation of calcium butyrate on growth performance, carcass traits, intestinal health, and pro-inflammatory cytokines in Japanese quails. *Journal of animal physiology and animal nutrition*, *103*(6), 1768-1775.
- 2. Abdelqader, A., and Al-Fataftah, A. R. (2016).Effect of dietary butyric acid on performance, intestinal morphology, microflora composition and intestinal recovery of heat-stressed broilers. *Livestock Science*, *183*, 78-83.
- 3. Ahsan, U., Cengiz, Ö., Raza, I., Kuter, E., Chacher, M. F. A., Iqbal, Z., and Çakir, S. (2016). Sodium butyrate in chicken nutrition: the dynamics of performance, gut microbiota, gut morphology, and immunity. *World's Poultry Science Journal*, *72*(2), 265-275.
- 4. Al-Gharban, H.A.A.J. (2017). Seroepidemiological detection and culture utilization for diagnosis of carrier horses and donkeys with strangles. *Journal of Education College Wasit University*, 1(28), 649-660.
- 5. Canani, R. B., Di Costanzo, M., Leone, L., Pedata, M., Meli, R., and Calignano, A. (2011). Potential beneficial effects of butyrate in intestinal and extraintestinal diseases. *World journal of gastroenterology: WJG*, *17*(12), 1519.
- 6. Castanon, J. I. R. (2007). History of the use of antibiotic as growth promoters in European poultry feeds. *Poultry science*, 86(11), 2466-2471.
- 7. Coban, H. B. (2020). Organic acids as antimicrobial food agents: Applications and microbial productions. *Bioprocess and Biosystems Engineering*, 43(4), 569-591.
- 8. Elnesr, S. S., Ropy, A., and Abdel-Razik, A. H. (2019).Effect of dietary sodium butyrate supplementation on growth, blood biochemistry, haematology and histomorphometry of intestine and immune organs of Japanese quail. *Animal*, *13*(6), 1234-1244.
- 9. Hajati, H., and Rezaei, M. (2010). The application of prebiotics in poultry production. *Int J Poult Sci*, 9(3), 298-304.

- 10. Hassan, H. M. A., Mohamed, M. A., Youssef, A. W., and Hassan, E. R. (2010).Effect of using organic acids to substitute antibiotic growth promoters on performance and intestinal microflora of broilers. *Asian-Australasian Journal of Animal Sciences*, 23(10), 1348-1353.
- 11. Hodin, R. (2000). Maintaining gut homeostasis: The butyrate–NF-κB connection. *Gastroenterology*, 118(4), 798-801.
- 12. Hu, Y. S., Guo, Y. G., Dominko, R., Gaberscek, M., Jamnik, J., and Maier, J. (2007). Improved electrode performance of porous LiFePO4 using RuO2 as an oxidic nanoscale interconnect. *Advanced Materials*, *19*(15), 1963-1966.
- 13. Jung, T. H., Park, J. H., Jeon, W. M., and Han, K. S. (2015). Butyrate modulates bacterial adherence on LS174T human colorectal cells by stimulating mucin secretion and MAPK signaling pathway. *Nutrition Research and Practice*, *9*(4), 343-349.
- 14. Khan, S. H., and Iqbal, J. (2016). Recent advances in the role of organic acids in poultry nutrition. *Journal of applied animal research*, 44(1), 359-369.
- 15. Kinoshita, M., Suzuki, Y., and Saito, Y. (2002). Butyrate reduces colonic paracellular permeability by enhancing PPARγ activation. *Biochemical and biophysical research communications*, 293(2), 827-831.
- 16. Leeson, S., Namkung, H., Antongiovanni, M., and Lee, E. H. (2005). Effect of butyric acid on the performance and carcass yield of broiler chickens. *Poultry science*, *84*(9), 1418-1422.
- 17. Lutful Kabir, S. M. (2009). The role of probiotics in the poultry industry. *International Journal of Molecular Sciences*, 10(8), 3531-3546.
- 18. Martínez, E. A., Babot, J. D., Lorenzo-Pisarello, M. J., Apella, M. C., and Chaia, A. P. (2016). Feed supplementation with avian Propionibacterium acidipropionici contributes to mucosa development in early stages of rearing broiler chickens. *Beneficial Microbes*, 7(5), 687-698.
- 19. Nagy, A. S.2007. Poultry Health Administration, Al Noor Publishing House.
- 20. National Research Council (NRC). 1994. Nutrient Requirements of Poultry. 9th rev. ed. National Academy Press, Washington.
- 21. Sikandar, A., Zaneb, H., Younus, M., Masood, S., Aslam, A., Ashraf, S., and Rehman, H. (2017). Protective effect of sodium butyrate on growth performance, immune responses, and gut mucosal morphometry in Salmonella-challenged broiler chickens. *International Journal of Agriculture and Biology*, 19, 1387-1393.
- 22. Singer, R. S., and Hofacre, C. L. (2006). Potential impacts of antibiotic use in poultry production. *Avian diseases*, 50(2), 161-172.
- 23. Tokić, V., Lazarević, M., Sinovec, Z., and Tokić, A. (2007). The influence of different feed additives to performances and immune response in broiler chicken. *Acta Veterinaria-Beograd*, *57*(2-3), 217-229.
- 24. Warnecke, T., and Gill, R. T. (2005). Organic acid toxicity, tolerance, and production in Escherichia coli biorefining applications. *Microbial cell factories*, *4*(1), 1-8.
- 25. Yang, Y., Iji, P. A., and Choct, M. (2009). Dietary modulation of gut microflora in broiler chickens: a review of the role of six kinds of alternatives to in-feed antibiotics. *World's Poultry Science Journal*, 65(1), 97-114.
- 26. Zhou, D. A., Pan, Q., Xin, F. Z., Zhang, R. N., He, C. X., Chen, G. Y., and Fan, J. G. (2017). Sodium butyrate attenuates high-fat diet-induced steatohepatitis in mice by improving gut microbiota and gastrointestinal barrier. *World journal of gastroenterology*, 23(1), 60.
- 27. Zhou, Z. Y., Packialakshmi, B., Makkar, S. K., Dridi, S., and Rath, N. C. (2014). Effect of butyrate on immune response of a chicken macrophage cell line. *Veterinary Immunology and Immunopathology*, *162*(1-2), 24-32.