

Review Article: Impact of *Leishmania* spp. on Public Health

Gufran J. Shamkhi ^{1*}, and Sheama Alali ²

¹ Department of Basic Science, College of Density, University of Wasit, Wasit, Iraq

² Department of Microbiology, College of Medicine, University of Wasit, Wasit, Iraq

Email: gaeeli@uowasit.edu.iq ¹, shhamid@uowasit.edu.iq ²

* Corresponding author

Abstract

The genus of *Leishmania*, intracellular protozoan parasite, *Leishmania* that belongs to the Trypanosomatida Order under Kinetoplastea Class of Euglenozoa Phylum, includes a large number of species that infect a wide variety of vertebrates and invertebrates, such as rodents and humans resulting in variable clinical manifestations. Taxonomy of the parasite is not fully elucidated, and debate continues not only over the number of species in the subgenus, but also over the definition of species. Leishmaniasis, caused by several different strains of *Leishmania*, remains a major public health problem globally. The disease is transmitted by mosquitoes and is endemic in mainly arthropod-carrying tropical areas. Due to fast changes in global climate, mosquitoes are expected to expand widely to become more susceptible to disease. There are four main types of disease including cutaneous leishmaniasis (CL), diffuse cutaneous (DCL), mucocutaneous leishmaniasis (MCL) and visceral leishmaniasis (VL) which varies in their characteristics. However, sequelae of the disease based upon the species of parasite and an immune response. With few treatments available, antimonials compounds remain very important in the therapeutic strategies which used most commonly in different areas. However, drug resistance has recently emerged including resistance to the oral therapy for the most recent drug used against VL. This alarming has led to use of non-toxic drugs to treatment of disease and to prescription-related complications. The prospect of widespread drug resistance therefore indicates an urgent need to develop effective new treatments for leishmaniasis. In conclusion, there is a necessary for limiting the complications and preventing death. Additionally, vector control can be implemented wherever possible.

Keywords: Cutaneous leishmaniasis, Diffuse-cutaneous leishmaniasis, Mucocutaneous leishmaniasis, Visceral leishmaniasis, Immunotherapy, Iraq

1.1. Introduction

Leishmaniasis is a disease that caused by several different strains of the intracellular protozoan parasite, *Leishmania* that belongs to the Trypanosomatida Order under Kinetoplastea Class of Euglenozoa Phylum. This parasite remains a major public health problem globally in four ecoepidemiological regions (Ruiz-Postigo et al., 2021). The disease is transmitted by mosquitoes and is endemic in mainly arthropod-carrying tropical areas. As global climate change continues, mosquitoes are expected to expand in range to become more susceptible to disease (Amro et al., 2022). The parasite is spread by sand-flies that carry flagellated promastigotes, and during feeding, the mosquito injects promastigotes into the skin to be engulfed rapidly by the phagocytic cells. Although, infected cells include neutrophils, macrophages and monocytes, but these organisms reside primarily in macrophages and manifest as infectious amastigotes (Teixeira et al., 2013). Surprisingly, unlike other intracellular parasites such as *Toxoplasma gondii* and *Trypanosoma cruzi*, *Leishmania* has evolved the ability to survive, multiply in phagolysosomes, and survive in an environment capable of killing many bacteria (Kolářová and Valigurová, 2021). After several rounds of replication, amastigotes infect infected cells and invade other nearby macrophages. When taken up by the sand-flies, the protozoan divides into promastigotes and the infectious cycle (Van Assche et al., 2011). Due to the geographical distribution of different species, *Leishmania* protozoa causes mainly three types of disease are cutaneous (KL) and mucocutaneous (MCL) and visceral (VL) leishmaniasis which occurs as a result of infection of macrophages throughout the skin, nasopharyngeal mucosa, or endothelial reticular system, respectively, and the sequelae of disease is based upon the species of parasite and the immune response (Van Griensven et al., 2014).

Although, there are many types of *Leishmania* such as *L. amazonensis*, *L. brazili*, *L. ethiopia*, *L. guayanensis*, *L. major*, *L. mexicana*, *L. panamensis*, *L. peruviana*, and *L. tropica*; few treatments available, antimonials compounds remain very important in the therapeutic strategies used in the most commonly used areas (Mohammadiha et al., 2018). Despite resistance in some *Leishmania* species, initial treatment of leishmaniasis lasted approximately 70 years (Mohabali et al., 2019). Unfortunately, there has been no vaccine for leishmaniasis for many years and disease control relies primarily on chemotherapy (Olías-Molero et al., 2021). The first line of treatment is a pentavalent meglumine antimonate (glucanthyme) indicated for the treatment of leishmaniasis. The current second line of treatment is sodium stibogluconate. Pentostam is a short course of amphotericin B containing pentamidine isothionate and miltefosine and is used when first-line drugs do not respond adequately (Seifert et al., 2011). Use of these substances is injectable and can be very dangerous. Furthermore, drug resistance has recently emerged in *Leishmania*, including resistance to the oral alkylphospholipid miltefosine, the most recent drug used against VL (Olivier and Zamboni, 2020). This alarming trend has led to the use of non-toxic drugs to treat the disease and/or subject to prescription-related complications. The prospect of widespread drug resistance therefore indicates an urgent need to develop effective new treatments for leishmaniasis (Carvalho et al., 2019). Hence, the current study was aimed to focus on the worldwide distribution of leishmaniasis.

1.2. Taxonomic classification

The genus of *Leishmania* includes a large number of species demonstrating that this parasite can utilize a wide variety of vertebrates and invertebrates, such as rodents and humans, as an intermediate host and to cause a wide range of clinical manifestations (Akhoundi et al., 2016). Taxonomy is not fully elucidated, and debate continues not only over the number of species in the subgenus, but also over the definition of species (Cecílio et al., 2022). Recent molecular data and phylogenetic analyzes support the simplification of *Leishmania* taxonomy to multiple species (Maurício, 2018).

1.3. Life Cycle

Leishmania lives as an intracellular parasite (amastigote) in macrophages of mammals and other vertebrates, and as an extracellular parasite (promastigote) in the gut of insect vectors (De Muylder et al., 2011; Kelly et al., 2017). The insect absorbs the blood of its vertebrate host and regurgitates the promastigotes upon skin infection. Parasites are recognized and phagocytosed by surface receptors on macrophages and dendritic cells. Interestingly, the parasite has a hard time infecting (and surviving) neutrophil-deficient animal. This result strongly suggests the relevance of a macrophage invasion mechanism that uses polymorphonuclear leukocytes as the first phagocytic cells encountered within the host. The parasite induces programmed cell death in infected neutrophils and is recruited by macrophages (Liu and Uzonna, 2012; von Stebut and Tenzer, 2018; Kupani et al., 2021). Within host cell, the parasite moves into phagolysosomes, divides into amastigotes, and multiplies by binary fission (Shirbazou and Jafari, 2012). Ablation of macrophages releases infected amastigotes, and released parasites are engulfed by simple macrophages, dramatically increasing the number of infected cells and spreading the disease throughout the host (Dar et al., 2018). Feeding on blood-sucking insects of infected individuals, the amastigotes develop into promastigotes in the digestive tract of the vector insect, where they survive for 4-7 days, differentiate into infected parasites, migrate to heart valves, and promastigote. When the mosquito re-pierces the host's skin, the bacteria enter the bloodstream and complete the cycle (Telleria et al., 2012; Teixeira et al., 2013; Inbar et al., 2017).

1.4. Vector

Mosquitoes are part of the family *Psychodidae*, a subfamily of *Phlebotomina* (Akhoundi et al., 2016). The four stages are the adult aerial stage and the stage of development from the egg, the larvae and pupae found in moist soils rich in organic matter (Solano-Gallego et al., 2017). Adults are small flying insects, about 2 to 4 mm long, with yellow hairs on their bodies (Claborn, 2010). They hide in the dark during the day and act in the dark and at night. Both sexes eat plants, but females also need blood before ovulation of reptiles, amphibians, birds, and mammals that act as the main hosts (Klein and Flanagan, 2016). Feeding habits depend on the mosquito species and host conditions, and the main factor in the transmission of leishmaniasis is the blood meal that mosquitoes ingest (Pruzinova et al., 2015). About 800 species have been described and classified into five widely accepted genera. Of these, approximately 70 species in the genera *Phlebotomy* and *Lutzomyia* are known or suspected hosts of *Leishmania*, with some specificity between *Leishmania* species and mosquitoes. Severe infections are mainly caused by his *Phlebotomus papatasi*. The

housefly *Phlebotomus sergenti* is the primary vector for *L. tropica* in Middle East (Dostálová and Volf, 2012; Lahouiti et al., 2013; Khan and Awan, 2021).

1.5. Origin

Most species of *Leishmania* are animals, with various mammalian reservoirs involved in the long-term maintenance of *Leishmania* in nature (Alemayehu and Alemayehu, 2017). Depending on host, reservoirs can be wild or domesticated mammals, possibly humans. Most leishmaniasis reservoirs are well-adapted and develop mild disease that can last for years, with the exception of dogs, which often develop chronic fatal disease. These instruments are available in 7 different styles for mammals (Alemayehu and Alemayehu, 2017; Abbate et al., 2019). Mice, hyraxes, marsupials, and carnivores are common enemies of cutaneous leishmaniasis in humans and wild animals. Humans have reservoirs of *L. donovani* and *L. tropica*. The locality of *Rhombomys opimus* is a major reservoir of important in the arid regions (Medkour et al., 2019; Pareyn et al., 2020).

1.6. Types of disease

Leishmaniasis causes significant morbidity and mortality (Al-Kamel, 2017). A common term for several diseases with different clinical manifestations, leishmaniasis is divided into four main types by him based on clinical manifestations. The fatal form that may leave without treatment is visceral leishmaniasis (VL). There are many other forms of CL and MCL which cause significant morbidity in endemic settings (Banerjee et al., 2016; Sunter and Gull, 2017).

1.6.1. CL

This is the most common form of disease, which manifested by a simple skin lesion appears at site of mosquito biting and heals within a few months, leaving a scar (Remadi et al., 2016). Incubation could be lasted to several weeks, and gradually enlarges and becomes red. Wound healing involves migration of leukocytes that divide the affected area, resulting in necrosis of the affected tissue and healing of granulomas. Bark lesions are the most common clinical manifestation of *L. majoris* infection, occurring mainly on the extremities (Iqbal, 2012; Aoun et al., 2014). Wounds become dangerously infected, multiple ulcers and moist mucus are common, and healing is slow. Self-healing occurs within 2 to 8 months in more than half of patients (Keogan, 2018). It can also appear as large dry, scaly, or ulcerated form often appears on the face and other exposed areas of *L. tropica* patients (Abdellatif et al., 2013; Rostamian and Niknam, 2019).

1.6.2. DCL

It is chronic, persistent and polyparasitic species occurs with certain forms of leishmaniasis and leprosy-like non-ulcerative skin lesions. The main pathogens are *L. aethiopia* and *L. mexicana* species complex (Tabah, 2018; Guma, 2018; Malli et al., 2019).

1.6.3. MCL

This type causes horrific ulcers, facial cuts, and severe destruction of the nasal, oropharyngeal, and pharyngeal cavities with chronic pain. MCL has been reported occasionally in Sudan (Babiker et al., 2014; Carvalho et al., 2019). MCL usually exists as an unmanned disease whose life cycle transitions from rodent to rodent and mammalian and is spread by the forest floor mosquito, *Lutzumia* spp. (Silva-Almeida et al., 2012; Borghi et al., 2017).

1.6.4. VL

It is highly dangerous and destructive, which also named as kalaazar, black disease, black fever, buldwan fever, dum dum fever, and sarkari disease (Pradhan et al., 2022). The organism is responsible for many clinical manifestations that can progress from asymptomatic infections to the fatal form of VL in severe cases (Chakravarty et al., 2019; Lewis et al., 2020; Mann et al., 2021).

1.7. Diagnosis

Diagnosis of CL is based on clinical features, epidemiological data, and laboratory tests. Traditionally, diagnostic tests for CL are classified as direct tests, including microscopic parasitological examination, histology and parasite culture (visual observation of *Leishmania* parasites), or indirect tests, including serology and molecular diagnostics. It's been done. The choice of which diagnostic test is used depends on diagnostic center infrastructure and capacity rather than diagnostic accuracy (de Vries et al., 2015; Handler et al., 2015; Reimão et al., 2020). Accurate identification of Giemsa-stained lesion scrape astigmatism by sampling, scraping, or imaging is frequently used to confirm the diagnosis in routine cases and in human

participants of clinical trials (Pourmohammadi et al., 2010). Parasite culture is a laborious and time-consuming procedure typically performed in reference centers and laboratories (Rasti et al., 2016). Several serological tests have been tested, but only a few are commercially available and used, possibly due to low sensitivity, poor testicular response to infection, or difficulty in obtaining a positive test. Serological testing is primarily based on enzyme-linked immunosorbent assay (ELISA), Western blot, lateral flow assay, or direct correlation (Lévêque et al., 2020; Piyasiri et al., 2022). Molecular diagnostic assays appear to have better sensitivity and specificity than traditional diagnostic methods, requiring less sampling for diagnosis. Molecular assays also allow identification of *Leishmania* species that infect patients. This is important change that contains useful data for evaluating clinical trials and selecting the best treatment for infected patients. PCR has been extensively evaluated as a single assay, in a nested format, or as a quantitative assay, using basic principles and sources of clinical material (Saldarriaga et al., 2016; Sundar and Singh, 2018; Schallig et al., 2019). Clinical manifestations, especially regeneration of ulcerative lesions or flattening of un-respectable wounds, are used as parameters to determine the clinical treatment of CL (Kassiri et al., 2014). Because the presence of parasites in healing wounds, association with clinical recurrence is unclear, most clinical trials measure treatment outcomes based solely on clinical observations (Simon et al., 2011).

1.8. Treatment

1.8.1. Pentavalent antimony (Sbv)

In India, sodium stibogluconate (Pentostam®, GlaxoSmithKline and generics) and meglumine antimonate (AM) (Glucanthyme™, Sanofi), commonly known as SSG, have been used in LC since their discovery by Albert David in the 1940s has been used as a first-line treatment for came out (Moore and Lockwood, 2010; Patino et al., 2019). The mechanism the drug by which act is not fully understood; however when used, Sbv (SbIII) is an active secondary metabolite, which inhibits DNA topoisomerases and causes depletion of intracellular ATP, possibly by inhibiting glycolysis and β -fatty acid oxidation in amastigotes (Keshav et al., 2021). This inhibits amastigote macromolecular biosynthesis (Fatima et al., 2016). Pentavalent antihistamines are prescribed for local or systemic treatments. In patients with more complicated CL, anticonvulsants can be administered systemically (Cuestas et al., 2018).

1.8.2. Amphotericin B

The polyene antibiotic amphotericin B (AmB) is commonly prescribed for VL and CL treatment (Solomon et al., 2011; Brotherton et al., 2014). Common formulations of drug cause severe nephrotoxicity that lowers lipid levels (Sundar et al., 2015). It acts through modulating of parasite membrane permeability by activating trans-membrane channels, causing a decrease in parasite ionic levels and leading to parasite death (Shirzadi, 2019).

1.8.3. Miltefosine

Miltefosine is a systemic drug for VL and CL cure, which exhibits highly time-dependent effects in pharmacokinetic and pharmacodynamic studies significant activity against various *Leishmania* species (Dorlo et al., 2012). This activity against *Leishmania* is associated with impaired alkylphospholipid metabolism and glycolipid and glycoprotein biosynthesis in the parasite's outer membrane (Rios-Marco et al., 2017). Another postulated mechanism is cell-mediated activation of macrophages to undergo apoptosis via the phosphoinositide-3-kinase pathway (Crauwels et al., 2015; Palić et al., 2019). The drug might be a teratogenic and should be avoided in women of childbearing (Vakil et al., 2015).

1.8.4. Paromomycin Sulphate

It is aminoglycoside antibiotic acting through binding to small subunit of ribosome, causing misfolding and inhibition of bacterial protein synthesis. It was found to have antipsychotic properties in the 1960s (de Morais-Teixeira et al., 2014; Santos et al., 2020). Another study suggested that it may cause mistranslations, thereby compromising the accuracy of *Leishmania* protein synthesis (Kattoof, 2018). Elevated levels of RNA misread result in a host growth arrest properties of parasites (Oliveira et al., 2021).

1.8.5. Topical treatment

Evaluation of local therapy is also very difficult because the dose is difficult to measure during administration. Topical therapy is of high activity but the skin barrier can be a problem. Local injection of drugs is most reliable treatment for CL and several studies have compared treatment with injection in experimentally treatments (Solomon et al., 2014; Azim et al., 2021). A meta-analysis conducted and the placebo-control trials of topical treatment were therapeutically effective in new and old CL and increased

local reactions (Cota et al., 2016). Many topical agents are compound preparations used for systemic therapy and are prescribed as cream or ointment with high antiparasitic activity and lack of systemic toxicity when applied to CL lesions (Carneiro et al., 2012; Escrivani et al., 2020; Jamshaid et al., 2021).

1.8.6. Physiotherapy

Cryotherapy and heat therapy have commonly methods of physical therapy which applied based on extreme temperatures to kill a parasite without causing significant damage to the host and the body can repair damage to healthy skin. Cryotherapy consists of applying liquid nitrogen-filled potatoes for 10-25 seconds (Jowkar et al., 2012; Shaddel et al., 2018). There are different uses of this method including the application of Cryotherapy alone, cryotherapy plus intralesional meglumine antimonate (MA), and intralesional MA alone (Ullah et al., 2022). Complete cure in these methods as reported by different studies was ranged 52-67% for cryotherapy alone, 80-89% for cryotherapy and intralesional MA, and 52-75% for intralesional MA alone (Brito et al., 2017; Ullah et al., 2022).

1.8.7. Immunotherapy

Unlike chemotherapy, this type of treatment can stimulate immune response for destroying of parasite through production of nitric oxide (NO) in macrophages, improving infection-fighting (Ikeogu et al., 2020; Akbari et al., 2021). Host protective effects were also associated with reduced ear parasite burden and lymph node recruitment. Topical administration of immune-therapy can reduce disease severity, and protected against both resistant and susceptible hosts (Adhikari et al., 2012; Montakhab-Yeganeh et al., 2017).

1.9. Control strategy

Effective treatment depends on earlier detection and treatment. This necessary is for limiting the complications and preventing death (Mahmoud, 2014). Patient care is an effective means of managing the storage and transportation of human sites. Additionally, vector control can be implemented wherever possible. Household pesticide residue application was significant step with restriction used today (Romero and Boelaert, 2010; Werneck, 2014; Dantas-Torres et al., 2019).

Conclusion

Although, *Leishmania* is one of the most prevalent parasitic pathogen in the world, many aspects are still unknown and need to be clarified. However, the development the methods of control and prevention are of great importance to avoid the diseases and its complications.

References

1. Abbate, J. M., Arfuso, F., Napoli, E., Gaglio, G., Giannetto, S., Latrofa, M. S., and Brianti, E. (2019). *Leishmania infantum* in wild animals in endemic areas of southern Italy. *Comparative Immunology, Microbiology and Infectious Diseases*, 67, 101374.
2. Abdellatif, M. Z., El-Mabrouk, K., and Ewis, A. A. (2013). An epidemiological study of cutaneous leishmaniasis in Al-jabal Al-gharbi, Libya. *The Korean journal of parasitology*, 51(1), 75.
3. Adhikari, A., Gupta, G., Majumder, S., Banerjee, S., Bhattacharjee, S., Bhattacharya, P., and Majumdar, S. (2012). *Mycobacterium indicus pranii* (Mw) re-establishes host protective immune response in *Leishmania donovani* infected macrophages: critical role of IL-12. *PLoS One*, 7(7), e40265.
4. Akbari, M., Oryan, A., and Hatam, G. (2021). Immunotherapy in treatment of leishmaniasis. *Immunology Letters*, 233, 80-86.
5. Akhoundi, M., Kuhls, K., Cannet, A., Votýpka, J., Marty, P., Delaunay, P., and Sereno, D. (2016). A historical overview of the classification, evolution, and dispersion of *Leishmania* parasites and sandflies. *PLoS neglected tropical diseases*, 10(3), e0004349.
6. Alemayehu, B., and Alemayehu, M. (2017). Leishmaniasis: a review on parasite, vector and reservoir host. *Health Science Journal*, 11(4), 1.
7. Al-Kamel, M. A. N. (2017). Leishmaniasis and malignancy: A review and perspective. *Clinical Skin Cancer*, 2(1-2), 54-58.
8. Amro, A., Moskalenko, O., Hamarsheh, O., and Frohme, M. (2022). Spatiotemporal analysis of cutaneous leishmaniasis in Palestine and foresight study by projections modelling until 2060 based on climate change prediction. *Plos one*, 17(6), e0268264.

9. Aoun, J., Habib, R., Charaffeddine, K., Taraif, S., Loya, A., and Khalifeh, I. (2014). Caseating granulomas in cutaneous leishmaniasis. *PLoS neglected tropical diseases*, 8(10), e3255.
10. Azim, M., Khan, S. A., Ullah, S., Ullah, S., and Anjum, S. I. (2021). Therapeutic advances in the topical treatment of cutaneous leishmaniasis: A review. *PLoS Neglected Tropical Diseases*, 15(3), e0009099.
11. Babiker, A. M., Ravagnan, S., Fusaro, A., Hassan, M. M., Bakheit, S. M., Mukhtar, M. M., and Capelli, G. (2014). Concomitant infection with *Leishmania donovani* and *L. major* in single ulcers of cutaneous leishmaniasis patients from Sudan. *Journal of Tropical Medicine*, 2014.
12. Banerjee, A., Bhattacharya, P., Joshi, A. B., Ismail, N., Dey, R., and Nakhasi, H. L. (2016). Role of pro-inflammatory cytokine IL-17 in *Leishmania* pathogenesis and in protective immunity by *Leishmania* vaccines. *Cellular immunology*, 309, 37-41.
13. Borghi, S. M., Fattori, V., Conchon-Costa, I., Pinge-Filho, P., Pavanelli, W. R., and Verri, W. A. (2017). *Leishmania* infection: painful or painless?. *Parasitology research*, 116, 465-475.
14. Brito, N. C., Rabello, A., and Cota, G. F. (2017). Efficacy of pentavalent antimoniate intralesional infiltration therapy for cutaneous leishmaniasis: A systematic review. *PloS one*, 12(9), e0184777.
15. Brotherton, M. C., Bourassa, S., Légaré, D., Poirier, G. G., Droit, A., and Ouellette, M. (2014). Quantitative proteomic analysis of amphotericin B resistance in *Leishmania infantum*. *International Journal for Parasitology: Drugs and Drug Resistance*, 4(2), 126-132.
16. Carneiro, G., Aguiar, M. G., Fernandes, A. P., and Ferreira, L. A. M. (2012). Drug delivery systems for the topical treatment of cutaneous leishmaniasis. *Expert opinion on drug delivery*, 9(9), 1083-1097.
17. Carvalho, S. H., Frézard, F., Pereira, N. P., Moura, A. S., Ramos, L. M., Carvalho, G. B., and Rocha, M. O. (2019). American tegumentary leishmaniasis in Brazil: a critical review of the current therapeutic approach with systemic meglumine antimoniate and short-term possibilities for an alternative treatment. *Tropical Medicine and International Health*, 24(4), 380-391.
18. Cecílio, P., Cordeiro-da-Silva, A., and Oliveira, F. (2022). Sand flies: Basic information on the vectors of leishmaniasis and their interactions with *Leishmania* parasites. *Communications biology*, 5(1), 305.
19. Chakravarty, J., Hasker, E., Kansal, S., Singh, O. P., Malaviya, P., Singh, A. K., and Sundar, S. (2019). Determinants for progression from asymptomatic infection to symptomatic visceral leishmaniasis: A cohort study. *PLoS neglected tropical diseases*, 13(3), e0007216.
20. Claborn, D. M. (2010). The biology and control of leishmaniasis vectors. *Journal of global infectious diseases*, 2(2), 127.
21. Cota, G. F., de Sousa, M. R., Fereguetti, T. O., Saleme, P. S., Alvarisa, T. K., and Rabello, A. (2016). The cure rate after placebo or no therapy in American cutaneous leishmaniasis: a systematic review and meta-analysis. *PloS one*, 11(2), e0149697.
22. Crauwels, P., Bohn, R., Thomas, M., Gottwalt, S., Jäckel, F., Krämer, S., and Zandbergen, G. V. (2015). Apoptotic-like *Leishmania* exploit the host's autophagy machinery to reduce T-cell-mediated parasite elimination. *Autophagy*, 11(2), 285-297.
23. Cuestas, D., Forero, Y., Galvis, I., Peñaranda, E., Cortes, C., Motta, A., and Puentes, J. (2018). Drug reaction with eosinophilia and systemic symptoms (DRESS) and multiple organ dysfunction syndrome (MODS): one more reason for a new effective treatment against leishmaniasis. *International Journal of Dermatology*, 57(11), 1304-1313.
24. Dantas-Torres, F., Miró, G., Bowman, D. D., Gradoni, L., and Otranto, D. (2019). Culling dogs for zoonotic visceral leishmaniasis control: the wind of change. *Trends in parasitology*, 35(2), 97-101.
25. Dar, M. J., Din, F. U., and Khan, G. M. (2018). Sodium stibogluconate loaded nano-deformable liposomes for topical treatment of leishmaniasis: macrophage as a target cell. *Drug delivery*, 25(1), 1595-1606.
26. de Morais-Teixeira, E., Gallupo, M. K., Rodrigues, L. F., Romanha, A. J., and Rabello, A. (2014). In vitro interaction between paromomycin sulphate and four drugs with leishmanicidal activity against three New World *Leishmania* species. *Journal of Antimicrobial Chemotherapy*, 69(1), 150-154.

27. De Muylder, G., Ang, K. K., Chen, S., Arkin, M. R., Engel, J. C., and McKerrow, J. H. (2011). A screen against *Leishmania* intracellular amastigotes: comparison to a promastigote screen and identification of a host cell-specific hit. *PLoS Neglected Tropical Diseases*, 5(7), e1253.
28. de Vries, H. J., Reedijk, S. H., and Schallig, H. D. (2015). Cutaneous leishmaniasis: recent developments in diagnosis and management. *American journal of clinical dermatology*, 16, 99-109.
29. Dorlo, T. P., Balasegaram, M., Beijnen, J. H., and de Vries, P. J. (2012). Miltefosine: a review of its pharmacology and therapeutic efficacy in the treatment of leishmaniasis. *Journal of Antimicrobial Chemotherapy*, 67(11), 2576-2597.
30. Dostálová, A., and Volf, P. (2012). *Leishmania* development in sand flies: parasite-vector interactions overview. *Parasites and vectors*, 5(1), 1-12.
31. Escrivani, D. O., Lopes, M. V., Poletto, F., Ferrarini, S. R., Sousa-Batista, A. J., Steel, P. G., and Rossi-Bergmann, B. (2020). Encapsulation in lipid-core nanocapsules improves topical treatment with the potent antileishmanial compound CH8. *Nanomedicine: Nanotechnology, Biology and Medicine*, 24, 102121.
32. Fatima, N., Muhammad, S. A., Mumtaz, A., Tariq, H., Shahzadi, I., Said, M. S., and Dawood, M. (2016). Fungal metabolites and Leishmaniasis: A review. *Br. J. Pharm. Res*, 12, 1-12.
33. Guma, G. T. (2018). *Visceral leishmaniasis in Ethiopia: Innate immune functions, biomarkers of cure and potential roles of cattle for transmission* (Doctoral dissertation, Universität zu Lübeck).
34. Handler, M. Z., Patel, P. A., Kapila, R., Al-Qubati, Y., and Schwartz, R. A. (2015). Cutaneous and mucocutaneous leishmaniasis: differential diagnosis, diagnosis, histopathology, and management. *Journal of the American Academy of Dermatology*, 73(6), 911-926.
35. Ikeogu, N. M., Akaluka, G. N., Edechi, C. A., Salako, E. S., Onyilagha, C., Barazandeh, A. F., and Uzonna, J. E. (2020). *Leishmania* immunity: advancing immunotherapy and vaccine development. *Microorganisms*, 8(8), 1201.
36. Inbar, E., Hughitt, V. K., Dillon, L. A., Ghosh, K., El-Sayed, N. M., and Sacks, D. L. (2017). The transcriptome of *Leishmania* major developmental stages in their natural sand fly vector. *MBio*, 8(2), e00029-17.
37. Iqbal, H. (2012). Comparative efficacy of *Aloe vera* and *Tamarix aphylla* against cutaneous leishmaniasis. *International Journal of Basic Medical Sciences and Pharmacy (IJBMS)*, 2(2).
38. Jamshaid, H., Din, F. U., and Khan, G. M. (2021). Nanotechnology based solutions for anti-leishmanial impediments: a detailed insight. *Journal of Nanobiotechnology*, 19, 1-51.
39. Jowkar, F., Dehghani, F., and Jamshidzadeh, A. (2012). Is topical nitric oxide and cryotherapy more effective than cryotherapy in the treatment of old world cutaneous leishmaniasis?. *Journal of dermatological treatment*, 23(2), 131-135.
40. Kassiri, H., Lotfi, M., Farajifard, P., and Kassiri, E. (2014). Laboratory diagnosis, clinical manifestations, epidemiological situation and public health importance of cutaneous leishmaniasis in Shushtar County, Southwestern Iran. *Journal of Acute Disease*, 3(2), 93-98.
41. Kattoof, W. M. (2018). Intralesional streptomycin: New, safe, and effective therapeutic option for cutaneous leishmaniasis. *Mustansiriyah Medical Journal*, 17(1), 42-46.
42. Kelly, P. H., Bahr, S. M., Serafim, T. D., Ajami, N. J., Petrosino, J. F., Meneses, C., and Wilson, M. E. (2017). The gut microbiome of the vector *Lutzomyia longipalpis* is essential for survival of *Leishmania infantum*. *MBio*, 8(1), e01121-16.
43. Keogan, D. (2018). *The Development of Metallohydroxamates as Novel Anti-Bacterial and Anti-Leishmanial Agents* (Doctoral dissertation, Royal College of Surgeons in Ireland).
44. Keshav, P., Goyal, D. K., and Kaur, S. (2021). Promastigotes of *Leishmania donovani* exhibited sensitivity towards the high altitudinal plant *Cicer microphyllum*. *Current Research in Parasitology and Vector-borne Diseases*, 1, 100040.
45. Khan, W., and Awan, Z. U. R. (2021). LEISHMANIA TROPICA A CUTANEOUS DISEASE DETECTED IN PEOPLE RESIDING IN NORTH WAZIRISTAN DISTRICT, KHYBER PAKHTUNKHWA PAKISTAN. *FUUAST Journal of Biology*, 11(2), 119-124.
46. Klein, S. L., and Flanagan, K. L. (2016). Sex differences in immune responses. *Nature Reviews Immunology*, 16(10), 626-638.

47. Kolářová, I., and Valigurová, A. (2021). Hide-and-Seek: A game played between parasitic protists and their hosts. *Microorganisms*, 9(12), 2434.
48. Kupani, M., Pandey, R. K., and Mehrotra, S. (2021). Neutrophils and Visceral Leishmaniasis: Impact on innate immune response and cross-talks with macrophages and dendritic cells. *Journal of Cellular Physiology*, 236(4), 2255-2267.
49. Lahouiti, K., Maniar, S., and Bekhti, K. (2013). Seasonal fluctuations of phlebotomines sand fly populations (Diptera: Psychodidae) in the Moulay Yacoub province, centre Morocco: effect of ecological factors. *African Journal of Environmental Science and Technology*, 7(11), 1028-1031.
50. Lévêque, M. F., Lachaud, L., Simon, L., Battery, E., Marty, P., and Pomares, C. (2020). Place of serology in the diagnosis of zoonotic leishmaniasis with a focus on visceral leishmaniasis due to *Leishmania infantum*. *Frontiers in Cellular and Infection Microbiology*, 10, 67.
51. Lewis, M. D., Paun, A., Romano, A., Langston, H., Langner, C. A., Moore, I. N., and Sacks, D. L. (2020). Fatal progression of experimental visceral leishmaniasis is associated with intestinal parasitism and secondary infection by commensal bacteria, and is delayed by antibiotic prophylaxis. *PLoS pathogens*, 16(4), e1008456.
52. Liu, D., and Uzonna, J. E. (2012). The early interaction of *Leishmania* with macrophages and dendritic cells and its influence on the host immune response. *Frontiers in cellular and infection microbiology*, 2, 83.
53. Mahmoud, M. Z. (2014). Assessment of visceral leishmaniasis consequences using ultrasound. *Open Journal of Radiology*, 2014.
54. Malli, S., Pomel, S., Ayadi, Y., Deloménie, C., Da Costa, A., Loiseau, P. M., and Bouchemal, K. (2019). Topically applied chitosan-coated poly (isobutylcyanoacrylate) nanoparticles are active against cutaneous leishmaniasis by accelerating lesion healing and reducing the parasitic load. *ACS Applied Bio Materials*, 2(6), 2573-2586.
55. Mann, S., Frasca, K., Scherrer, S., Henao-Martínez, A. F., Newman, S., Ramanan, P., and Suarez, J. A. (2021). A review of leishmaniasis: current knowledge and future directions. *Current tropical medicine reports*, 8, 121-132.
56. Maurício, I. L. (2018). *Leishmania* taxonomy. *The Leishmaniasis: Old Neglected Tropical Diseases*, 15-30.
57. Medkour, H., Davoust, B., Dulieu, F., Maurizi, L., Lamour, T., Marie, J. L., and Mediannikov, O. (2019). Potential animal reservoirs (dogs and bats) of human visceral leishmaniasis due to *Leishmania infantum* in French Guiana. *PLoS Neglected Tropical Diseases*, 13(6), e0007456.
58. Mohammadiha, A., Dalimi, A., Mohebbali, M., Sharifi, I., Mahmoudi, M., Mirzaei, A., and Ghorbanzadeh, B. (2018). Molecular identification and phylogenetic classification of *Leishmania* spp. isolated from human cutaneous leishmaniasis in Iran: A cross-sectional study. *Iranian journal of parasitology*, 13(3), 351.
59. Mohebbali, M., Kazemirad, E., Hajjarian, H., Kazemirad, E., Oshaghi, M. A., Raoofian, R., and Teimouri, A. (2019). Gene expression analysis of antimony resistance in *Leishmania tropica* using quantitative real-time PCR focused on genes involved in trypanothione metabolism and drug transport. *Archives of dermatological research*, 311, 9-17.
60. Montakhab-Yeganeh, H., Abdossamadi, Z., Zahedifard, F., Taslimi, Y., Badirzadeh, A., Saljoughian, N., and Rafati, S. (2017). *Leishmania tarentolae* expressing CXCL-10 as an efficient immunotherapy approach against *Leishmania major*-infected BALB/c mice. *Parasite Immunology*, 39(10), e12461.
61. Moore, E. M., and Lockwood, D. N. (2010). Treatment of visceral leishmaniasis. *Journal of global infectious diseases*, 2(2), 151.
62. Olías-Molero, A. I., de la Fuente, C., Cuquerella, M., Torrado, J. J., and Alunda, J. M. (2021). Antileishmanial drug discovery and development: time to reset the model?. *Microorganisms*, 9(12), 2500.
63. Oliveira, S. S., Ferreira, C. S., Branquinha, M. H., Santos, A. L., Chaud, M. V., Jain, S., and Severino, P. (2021). Overcoming multi-resistant leishmania treatment by nanoencapsulation of potent antimicrobials. *Journal of Chemical Technology and Biotechnology*, 96(8), 2123-2140.

64. Olivier, M., and Zamboni, D. S. (2020). Leishmania Viannia guyanensis, LRV1 virus and extracellular vesicles: a dangerous trio influencing the faith of immune response during mucocutaneous leishmaniasis. *Current Opinion in Immunology*, 66, 108-113.
65. Palić, S., Bhairosing, P., Beijnen, J. H., and Dorlo, T. P. (2019). Systematic review of host-mediated activity of miltefosine in leishmaniasis through immunomodulation. *Antimicrobial agents and chemotherapy*, 63(7), e02507-18.
66. Pareyn, M., Kochora, A., Van Rooy, L., Eligo, N., Vanden Broecke, B., Girma, N., and Massebo, F. (2020). Feeding behavior and activity of Phlebotomus pedifer and potential reservoir hosts of Leishmania aethiopica in southwestern Ethiopia. *PLoS neglected tropical diseases*, 14(3), e0007947.
67. Patino, L. H., Muskus, C., and Ramírez, J. D. (2019). Transcriptional responses of Leishmania (Leishmania) amazonensis in the presence of trivalent sodium stibogluconate. *Parasites and vectors*, 12(1), 1-15.
68. Piyasiri, S. B., Samaranayake, T. N., Silva, H., Manamperi, N. H., and Karunaweera, N. D. (2022). ELISA-based evaluation of antibody response to Leishmania in a region endemic for cutaneous leishmaniasis. *Parasite Immunology*, 44(9), e12940.
69. Pourmohammadi, B., Motazedian, M. H., Hatam, G. R., Kalantari, M., Habibi, P., and Sarkari, B. (2010). Comparison of three methods for diagnosis of cutaneous leishmaniasis.
70. Pradhan, S., Schwartz, R. A., Patil, A., Grabbe, S., and Goldust, M. (2022). Treatment options for leishmaniasis. *Clinical and experimental dermatology*, 47(3), 516-521.
71. Pruzinova, K., Sadlova, J., Seblova, V., Homola, M., Votycka, J., and Volf, P. (2015). Comparison of bloodmeal digestion and the peritrophic matrix in four sand fly species differing in susceptibility to Leishmania donovani. *PloS one*, 10(6), e0128203.
72. Rasti, S., Ghorbanzadeh, B., Kheirandish, F., Mousavi, S. G., Pirozmand, A., Hooshyar, H., and Abani, B. (2016). Comparison of molecular, microscopic, and culture methods for diagnosis of cutaneous leishmaniasis. *Journal of Clinical Laboratory Analysis*, 30(5), 610-615.
73. Reimão, J. Q., Coser, E. M., Lee, M. R., and Coelho, A. C. (2020). Laboratory diagnosis of cutaneous and visceral leishmaniasis: current and future methods. *Microorganisms*, 8(11), 1632.
74. Remadi, L., Haouas, N., Chaara, D., Slama, D., Chargui, N., Dabghi, R., and Babba, H. (2016). Clinical presentation of cutaneous leishmaniasis caused by Leishmania major. *Dermatology*, 232(6), 752-759.
75. Rios-Marco, P., Marco, C., Gálvez, X., Jiménez-López, J. M., and Carrasco, M. P. (2017). Alkylphospholipids: An update on molecular mechanisms and clinical relevance. *Biochimica et Biophysica Acta (BBA)-Biomembranes*, 1859(9), 1657-1667.
76. Romero, G. A., and Boelaert, M. (2010). Control of visceral leishmaniasis in Latin America—a systematic review. *PLoS neglected tropical diseases*, 4(1), e584.
77. Rostamian, M., and Niknam, H. M. (2019). Leishmania tropica: What we know from its experimental models. *Advances in Parasitology*, 104, 1-38.
78. Ruiz-Postigo, J. A., Jain, S., Mikhailov, A., Maia-Elkhoury, A. N., Valadas, S., Warusavithana, S., and Gasimov, E. (2021). Global leishmaniasis surveillance: 2019-2020, a baseline for the 2030 roadmap/Surveillance mondiale de la leishmaniose: 2019-2020, une periode de reference pour la feuille de route a l'horizon 2030. *Weekly epidemiological record*, 96(35), 401-420.
79. Saldarriaga, O. A., Castellanos-Gonzalez, A., Porrozzi, R., Baldeviano, G. C., Lescano, A. G., de Los Santos, M. B., and Travi, B. L. (2016). An innovative field-applicable molecular test to diagnose cutaneous Leishmania Viannia spp. infections. *PLoS neglected tropical diseases*, 10(4), e0004638.
80. Santos, S. S., de Araujo, R. V., Giarolla, J., El Seoud, O., and Ferreira, E. I. (2020). Searching for drugs for Chagas disease, leishmaniasis and schistosomiasis: a review. *International journal of antimicrobial agents*, 55(4), 105906.
81. Schallig, H. D., Hu, R. V., Kent, A. D., van Loenen, M., Menting, S., Picado, A., and Cruz, I. (2019). Evaluation of point of care tests for the diagnosis of cutaneous leishmaniasis in Suriname. *BMC infectious diseases*, 19(1), 1-6.

82. Seifert, K., Munday, J., Syeda, T., and Croft, S. L. (2011). In vitro interactions between sitamaquine and amphotericin B, sodium stibogluconate, miltefosine, paromomycin and pentamidine against *Leishmania donovani*. *Journal of antimicrobial chemotherapy*, 66(4), 850-854.
83. Shaddel, M., Sharifi, I., Karvar, M., Keyhani, A., and Baziar, Z. (2018). Cryotherapy of cutaneous leishmaniasis caused by *Leishmania major* in BALB/c mice: A comparative experimental study. *Journal of Vector Borne Diseases*, 55(1), 42.
84. Shirbazou, S., and Jafari, M. (2012). The multiple forms of *Leishmania major* in BALB/C mice lung in iran. *Iranian Journal of Parasitology*, 7(2), 99.
85. Shirzadi, M. R. (2019). Liposomal amphotericin B: a review of its properties, function, and use for treatment of cutaneous leishmaniasis. *Research and reports in tropical medicine*, 11-18.
86. Silva-Almeida, M., Pereira, B. A. S., Ribeiro-Guimarães, M. L., and Alves, C. R. (2012). Proteinases as virulence factors in *Leishmania* spp. infection in mammals. *Parasites and vectors*, 5, 1-10.
87. Simon, I., Wissing, K. M., Del Marmol, V., Antinori, S., Rimmelink, M., Nilufer Broeders, E., and Cascio, A. (2011). Recurrent leishmaniasis in kidney transplant recipients: report of 2 cases and systematic review of the literature. *Transplant Infectious Disease*, 13(4), 397-406.
88. Solano-Gallego, L., Cardoso, L., Pennisi, M. G., Petersen, C., Bourdeau, P., Oliva, G., and Baneth, G. (2017). Diagnostic challenges in the era of canine *Leishmania infantum* vaccines. *Trends in Parasitology*, 33(9), 706-717.
89. Solomon, M., Pavlotsky, F., Leshem, E., Ephros, M., Trau, H., and Schwartz, E. (2011). Liposomal amphotericin B treatment of cutaneous leishmaniasis due to *Leishmania tropica*. *Journal of the European Academy of Dermatology and Venereology*, 25(8), 973-977.
90. Solomon, M., Schwartz, E., Pavlotsky, F., Sakka, N., Barzilai, A., and Greenberger, S. (2014). *Leishmania tropica* in children: a retrospective study. *Journal of the American Academy of Dermatology*, 71(2), 271-277.
91. Sundar, S., and Singh, O. P. (2018). Molecular diagnosis of visceral leishmaniasis. *Molecular diagnosis and therapy*, 22(4), 443-457.
92. Sundar, S., Singh, A., Rai, M., and Chakravarty, J. (2015). Single-dose indigenous liposomal amphotericin B in the treatment of Indian visceral leishmaniasis: a phase 2 study. *The American journal of tropical medicine and hygiene*, 92(3), 513.
93. Sunter, J., and Gull, K. (2017). Shape, form, function and *Leishmania* pathogenicity: from textbook descriptions to biological understanding. *Open biology*, 7(9), 170165.
94. Tabah, E. N. (2018). *Skin neglected tropical diseases in Cameroon: the need for integrated control and elimination* (Doctoral dissertation, University_of_Basel).
95. Teixeira, D. E., Benchimol, M., Rodrigues, J. C., Crepaldi, P. H., Pimenta, P. F., and de Souza, W. (2013). The cell biology of *Leishmania*: how to teach using animations. *PLoS pathogens*, 9(10), e1003594.
96. Telleria, E. L., Sant'Anna, M. R., Ortigão-Farias, J. R., Pitaluga, A. N., Dillon, V. M., Bates, P. A., and Dillon, R. J. (2012). Caspar-like gene depletion reduces *Leishmania* infection in sand fly host *Lutzomyia longipalpis*. *Journal of Biological Chemistry*, 287(16), 12985-12993.
97. Ullah, N., Uzair, M., Khan, N. U., and Butt, G. (2022). Comparative cost-effectiveness of intralesional meglumine antimoniate alone versus cryotherapy plus intralesional meglumine antimoniate in cutaneous leishmaniasis: Experience from a high capacity dermatology centre. *Journal of Pakistan Association of Dermatologists*, 32(2), 353-359.
98. Vakil, N. H., Fujinami, N., and Shah, P. J. (2015). Pharmacotherapy for leishmaniasis in the United States: focus on miltefosine. *Pharmacotherapy: The Journal of Human Pharmacology and Drug Therapy*, 35(5), 536-545.
99. Van Assche, T., Deschacht, M., da Luz, R. A. I., Maes, L., and Cos, P. (2011). *Leishmania*-macrophage interactions: Insights into the redox biology. *Free Radical Biology and Medicine*, 51(2), 337-351.
100. Van Griensven, J., Carrillo, E., López-Vélez, R., Lynen, L., and Moreno, J. (2014). Leishmaniasis in immunosuppressed individuals. *Clinical Microbiology and Infection*, 20(4), 286-299.

101. von Stebut, E., and Tenzer, S. (2018). Cutaneous leishmaniasis: Distinct functions of dendritic cells and macrophages in the interaction of the host immune system with *Leishmania major*. *International Journal of Medical Microbiology*, 308(1), 206-214.
102. Werneck, G. L. (2014). Visceral leishmaniasis in Brazil: rationale and concerns related to reservoir control. *Revista de saude publica*, 48, 851-856.